

T4 DNA Ligase



Size:

Cat#.	Size
BG10201S	20000 U
BG10201L	100000 U

Concentration: 400 units/ μ L

Source: *E.coli* cells with a cloned T4 DNA ligase gene from bacteriophage T4.

Storage temperature: -20°C.

Exp.: 12 months(-15°C ~ -25°C)

Applications:

1. Cloning of restriction fragments
2. Cloning of PCR products
3. Joining of double stranded linkers and adapters to DNA
4. Nick repair in duplex DNA, RNA or DNA/RNA hybrids



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Description:

T4 DNA ligase catalyzes the formation of a phosphodiester bond between juxtaposed 5' phosphate and 3' hydroxyl termini in duplex DNA or RNA. With the 10 \times T4 DNA Ligase Buffer, the enzyme can ligate DNA sticky or blunt end fragments.

Reagent Supplied:

Cat#.	Size	Reagent	No.
BG10201S	20000 U	10 \times T4 DNA Ligase Reaction Buffer	1 mL
BG10201L	100000 U	10 \times T4 DNA Ligase Reaction Buffer	2 \times 1 mL

Quality Control:

1. Protein Purity (SDS-PAGE): T4 DNA Ligase has >99% pure judged by SDS-PAGE.
2. Non-Specific DNase Activity (16 h): The product is tested for non-specific nuclease degradation in a reaction containing a DNA substrate. After incubation for 16 h there is no detectable degradation of the DNA substrate as determined by agarose gel electrophoresis.
3. Endonuclease Activity (Nicking): The product is tested in a reaction containing a supercoiled DNA substrate. After incubation for 4 h, < 5% DNA converted to the nicked form is determined by agarose gel electrophoresis.
4. qPCR for DNA Contamination(*E.coli* Genome): A minimum of 5 μ L of T4 DNA Ligase is screened for the presence of *E.coli* genomic DNA using qPCR with Taqman Probe specific for *E.coli* 16S rRNA. The measured level of *E.coli* genomic DNA contamination is less than 10 copies of *E.coli* genome.

Definition of activity unit:

One unit of the enzyme is defined as the amount of enzyme required to reach 50% ligation of 100ng HindIII fragments of λ DNA in a total reaction volume of 20 μ L in 30 min at 16°C in 1 \times T4 DNA Ligase Buffer.

Storage buffer:

The enzyme is supplied in: 10 mM Tris-HCl (pH7.5 @ 25°C), 50 mM KCl, 0.1 mM EDTA, 1 mM DTT and 50% (v/v) glycerol.

Protocol:

1. Thaw the 10 \times T4 DNA Ligase Reaction Buffer at room temperature and vortex vigorously to dissolve any precipitated material before use.
2. A molar ratio of 3:1 insert: vector(10-100ng) is recommended for the Reaction of DNA inserts to vectors. To calculate the amount of insert for a 3:1 ratio of vector to insert, use the formula:

$$X \text{ ng of insert} = \frac{3 \times \text{insert size (in bp)} \times \text{amount of vector (in ng)}}{\text{vector size (in bp)}}$$

Add the following reagents to an autoclaved, 0.2 mL microcentrifuge tube:

Reagent	Amount
10 \times T4 DNA Ligase Reaction Buffer	2 μ L
Linearized vector DNA	10-100 ng
Insert DNA	variable
T4 DNA Ligase	1 μ L
Nuclease-Free water	to 20 μ L

3. Mix gently and centrifuge briefly.

4. Incubate at room temperature for 10 min or 16°C overnight for sticky ends. Incubate at room temperature for 2 h or 16°C overnight for blunt ends or single base overhang.
5. Use 2-5 μ L of the ligation mixture for transform or Store at -20°C.

Reference:

Remaut, E., Tsao, H., Fiers, W., Improved plasmid vectors with a thermoinducible expression and temperature-regulated runaway replication [J]. *Gene*, 1983, 22(1) :103-113.

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T4 DNA 连接酶使用说明书

【产品名称】T4 DNA 连接酶

【产品规格】

货号	规格
BG10201S	20000 U
BG10201L	100000 U

【产品浓度】400 units/μL

【产品来源】重组表达 T4 噬菌体中 T4 DNA Ligase 基因的大肠杆菌

【储存条件】-20℃

【保质期】12 个月（-15℃~ -25℃）

【应用范围】

1. 限制性酶切片段的克隆
2. PCR 产物克隆
3. 将双链 linkers 或 adapters 连接到 DNA 上
4. dsDNA、dsRNA 或 DNA/RNA 杂交链的切刻修复

【产品说明】

T4 DNA 连接酶催化双链的 DNA 或 RNA 中相邻的 5'-磷酸基团和 3' 羟基末端形成磷酸二酯键。在含有 10×T4 DNA 反应缓冲液的环境下，T4 DNA 连接酶可以链接 DNA 的粘性末端和平末端。

【随酶提供试剂】

货号	规格	试剂	数量
BG10201S	20000 U	10×T4 DNA 反应缓冲液	1 mL
BG10201L	100000 U	10×T4 DNA 反应缓冲液	2×1 mL

【质量控制】

1. 蛋白纯度（SDS-PAGE）：SDS-PAGE 检测纯度 >99%
2. 非特异性核酸酶活性：将 T4 DNA 连接酶和 DNA 底物混合孵育 16 个小时，所得产物经过凝胶电泳未检测到 DNA 降解。

3. 内切酶（切刻）活性：将 T4 DNA 连接酶与超螺旋结构的 DNA 混合进行孵育 4h，< 5% 的超螺旋 DNA 转化为有切刻形式的 DNA。

【核酸污染】qPCR 检测 DNA 污染（大肠杆菌基因组）：通过 Taqman 探针 qPCR 法检测大肠杆菌 16S rRNA 位点，用至少 5 μL 的 T4 DNA 连接酶来检测，结果显示大肠杆菌基因组 DNA 的污染水平小于 10 个大肠杆菌基因组。

【单位定义】1 个单位指在 16℃，20 μL T4 DNA 反应缓冲液反应缓冲液条件下，30 分钟能使 50% 的经 Hind III 消化的 100ng λ DNA 片段所需的酶量。

【贮存条件】10 mM Tris-HCl (pH 7.5 @ 25°C), 50 mM KCl, 0.1 mM EDTA, 1 mM DTT 和 50%(w/v) 甘油。

【操作步骤】

1. 将 10×T4 DNA 反应缓冲液放置于室温中，请在使用前充分混匀。
2. 将 10~100 ng 的线性化载体和大于等于 3 倍摩尔量的插入片段进行混合，建议使用公式：

$$X \text{ ng of insert} = \frac{3 \times \text{insert size (in bp)} \times \text{amount of vector (in ng)}}{\text{vector size (in bp)}}$$

按下表要求在 0.2 mL 离心管中加入各种试剂：
具体反应体系如下：

试剂	含量
10×T4 DNA 反应缓冲液	2 μL
线性 DNA 载体	10~100 ng
插入 DNA 片段	可变量
T4 DNA Ligase	1 μL
无核酸酶水	至 20 μL

3. 混合均匀并短暂离心。
4. 如果是粘性末端的连接则在 16℃反应过夜或是在室温反

应 10min，如果是平末端或是单碱基的粘性末端，则 16℃反应过夜或是在室温中反应 2h。

5. 取 2-5 μL 产物转化或者放置于 -20℃进行储存。

【参考文献】

Remaut, E., Tsao, H., Fiers, W., Improved plasmid vectors with a thermoinducible expression and temperature-regulated runaway replication [J]. *Gene*, 1983, 22(1) : 103-113.

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